



Public Health Institute Ostrava
Centre of Clinical Laboratories
Location 1 - Ostrava
Partyzánské náměstí 2633/7, Moravská Ostrava, 702 00 Ostrava
VAT: CZ71009396



TEST REPORT n. 71/2022/V

Customer:

Vzduchotechnik, s.r.o.
Tovární ulice 548
463 31 Chrastava

Order number: 34835**Date of delivery:** 16.11.2022**Reference number:** ZU/34835/2022**Identification of sample:**

Number of sample:

71/2022

Name of sampleⁱ:

Microbicidal and virucidal nanofiber filter
membrane type XC130

Manufacturerⁱ:

Vzduchotechnik, s.r.o.

Batch numberⁱ:

/

Manufacturing date/ Expiry dateⁱ:

/

Sample compositionⁱ:

nanofiber polymer membrane, functionalized
with a natural, harmless substance
refrigerator (no light)

Storage conditionsⁱ:

grey coloured fabric

Appearance of the sample:

3.2.2023

Date of delivery:

ČSN ISO 18184:2020

Test method:

6.2.2023 – 16.2.2023

Date(s) of tests (period of analysis):

Location of tests:

Location 1 - Ostrava

ⁱ Data provided by customer.



Test procedure:

Test method: ČSN ISO 18184:2020 Textiles – Determination of antiviral activity of textile products

| | |
|----------------------|---|
| Tested virus strain: | <i>Vaccinia virus, strain Modified Vaccinia virus Ankara (ATCC-VR-1508)</i> |
| Cell line: | BHK-21 (ATCC-CCL-10) |
| Growth medium: | DMEM + 10% FBS |
| Maintenance medium: | EMEM + 2% FBS |

| | |
|-----------------------------|--|
| Neutralizer: | maintenance medium |
| Control material: | nanofiber filter membrane supplied by the customer |
| Method of sterilization: | samples and control material delivered sterile |
| Sample size: | 0,200 ± 0,025 g |
| Volume of viral suspension: | 100 µl |
| Volume of neutralizer: | 10 ml |
| Special light source: | ordinary laboratory lighting |
| Contact time(s): | 1 hour, 2 hours, 24 hours |
| Testing temperature: | 25 °C ± 1 °C |
| Incubation temperature: | 37 °C ± 1 °C |
| Method of filtration: | not used |
| Titration method: | viral titration on monolayer cell culture on the microplates |
| Titers calculated by: | Spaerman – Kärber's method |

Description of the test:

Under aseptic conditions, samples of control material and test material weighing 0,200 ± 0,025 g were prepared. The samples were transferred to sterile tubes. A given volume of viral suspension was applied to both control and tested samples. The samples were cultured at 25 °C ± 1 °C and a relative humidity of 90 %. After 24 hours, 2 hours and 1 hour of incubation, a given amount of neutralising solution was added to each sample and the samples were shaken on a vortex. After that, a series of tenfold dilutions of this mixture was prepared. Each dilution was titrated on a single-layer cell culture on a microtitre plate.

Three samples of both control and test material were used to verify the cytotoxicity of the material. A volume of neutralising medium corresponding to the viral suspension on the previous samples was applied to the samples. The samples were incubated at 25 °C ± 1 °C and a relative humidity of 90 %. After a contact period of 24 hours, 2 hours and 1 hour, a given amount of neutralising solution was added to each sample and the samples were shaken vigorously on the vortex. After that, a series of tenfold dilutions of this mixture was prepared. Each dilution was titrated on a single-layer cell culture on a microtitre plate.

Three samples of both control and test material were used to check the effectiveness of suppression of the activity of the test sample. A given amount of maintenance medium was applied to the samples. Immediately after application, a given quantity of neutralising solution was added to each sample and the samples were shaken vigorously. 50 µl of viral suspension was added to 5 ml of this suspension. After 30 minutes of incubation, a series of tenfold dilutions of this mixture is prepared. After 30 minutes of incubation, a series of tenfold dilutions of this mixture is prepared. Each dilution was titrated on a single-layer cell culture on a microtitre plate.

The virus titer values of all tests were determined 6th day of cultivation and calculated according to the Spaerman-Kärber method. The titer value is expressed as Ig TCID₅₀ - negative logarithm of 50% end point (50% infectious dose of virus suspension or such dilution of virus suspension that induces CPE in 50% of cell culture units).



Test results:

The test results are shown in the tables. These results are the average of three determinations of each sample and are expressed as the average of the calculated logarithms of the virus titres. The value of antiviral activity (M_v) is given in logarithms, it is expressed as the difference between the virus titre on the test sample (V_c) and the virus titre on the control (V_a).

$$M_v = \lg(V_a) - \lg(V_c)$$

Table n.1: The results for Vaccinia virus, strain Modified Vaccinia virus Ankara

| Sample | Level of cytotoxicity | Ig TCID ₅₀ /ml 1 hour | Ig TCID ₅₀ /ml 2 hours | Ig TCID ₅₀ /ml 24 hours | Value of antiviral activity (M_v) 1 hour | Value of antiviral activity (M_v) 2 hours | Value of antiviral activity (M_v) 24 hours |
|--|-----------------------|----------------------------------|-----------------------------------|------------------------------------|--|---|--|
| Microbicidal and virucidal nanofiber filter membrane XC130 (V _c) | 1,20 | 4,49 | 3,78 | 1,49 | 1,21 | 1,92 | 3,50 |
| Control (V _a) | 1,20 | 5,70 | 5,70 | 4,99 | | | |

Table n.2: Verificacion of cell sensitivity to virus and the inactivation of antiviral activity for Vaccinia virus, strain Modified Vaccinia virus Ankara

| Ig TCID ₅₀ /ml Microbicidal and virucidal nanofiber filter membrane XC130 | Ig TCID ₅₀ /ml Control material | Δ g TCID ₅₀ /ml | Control valid (Δ < 1) |
|---|---|----------------------------|--------------------------|
| 5,96 | 5,96 | 0,00 | Yes |

Table n.3: Control of test homogeneity

| Contact time | Ig TCID ₅₀ /ml max | Ig TCID ₅₀ /ml min | Ig TCID ₅₀ /ml mean | max – min mean | Control valid (≤0,2) |
|--------------|-------------------------------|-------------------------------|--------------------------------|----------------|----------------------|
| 0 hour | 6,07 | 5,45 | 5,74 | 0,11 | Yes |
| 1 hour | 5,82 | 5,57 | 5,70 | 0,04 | Yes |
| 2 hours | 5,82 | 5,57 | 5,70 | 0,04 | Yes |
| 24 hours | 5,32 | 4,82 | 4,99 | 0,10 | Yes |

All conditions for a valid test were satisfied.

Prepared by: Mgr. Markéta Tietzová



Conclusion:

According to the ČSN ISO 18184:2020 - contact of tested sample **71/2022** of the material **Microbicidal and virucidal nanofiber filter membrane XC130** with virus *Vaccinia virus, strain Modified Vaccinia virus Ankara* caused:

- Reduction of virus titer by 1,21 lg after contact time 1 hour
- Reduction of virus titer by 1,92 lg after contact time 2 hours
- Reduction of virus titer by 3,50 lg after contact time 24 hours

By testing according to the ČSN ISO 18184:2020, the sample **71/2022** of the material **Microbicidal and virucidal nanofiber filter membrane XC130** demonstrate excellent antiviral effect against the tested virus *Vaccinia virus, strain Modified Vaccinia virus Ankara* after 24 hours of contact.

Conclusion prepared by: Mgr. Markéta Tietzová

In Ostrava 7.4.2023

Zdravotní ústav se sídlem v Ostravě
Centrum klinických laboratoří
Oddělení virologie
Laboratoř pro testování virucidního účinku
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Moravská Ostrava 702 00 Ostrava
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Authorized by: Mgr. Markéta Tietzová
Laboratory for testing virucidal activity

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END OF THE PROTOCOL